

Collective behavior in cancer cell populations

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In recent years the argument has been made that malignant tumors represent complex dynamic and self-organizing biosystems. Furthermore, there is increasing evidence that collective cell migration is common during invasion and metastasis of malignant tumors. Here, we argue that cancer systems may be capable of developing multicellular collective patterns that resemble evolved adaptive behavior known from other biological systems including collective sensing of environmental conditions and collective decision-making. We present a concept as to how these properties could arise in tumors and why the emergence of such swarm-like patterns would confer advantageous properties to the spatiotemporal expansion of tumors, and consequently, why understanding and ultimately targeting such collectivity should be of interest for basic and clinical cancer research alike.

Keywords: biosystems; cancer; collective behavior

Introduction

Collective behavior has long been observed in a number of biological systems, most notably in social insect colonies (Fig. 1A), bird flocks, and schools of fish.⁽¹⁾ The behavior of such systems is complex,⁽²⁾ which means that understanding the individual component in more detail does not necessarily explain the collective behavior of many individuals, and thus usually evokes Aristotle's quote in that "The whole is more than the sum of its parts." Understanding the population-level properties of large interacting systems, and how such properties arise from the individual components, is a fundamental problem in the biological sciences.^(3–6)

In recent years it has been proposed that cancer behaves as such an expanding multicellular biosystem^(7–9) that results from the complicated multi-step process of tumorigenesis.⁽¹⁰⁾ Malignant tumors are generally thought to operate with a

combination of fast on-site growth and a (currently) nearly unstoppable population of rogue, mobile cancerous cells that invade the adjacent tissue and, depending on the cancer type, can eventually metastasize to other organs and set up secondary tumors.⁽¹¹⁾ In some microscopy images (such as the one in Fig. 1B taken from 2D cell culture experiments with highly malignant brain cancer cells), a pattern of collective alignment can be observed that seems to arise without any visible directional cue—not unlike the ones known from swarms (Fig. 1A). Such collective cell migration is now becoming recognized as being an important, and often the predominant mode of invasion in a wide range of tumors.^(12–14) Here, we argue that, not unlike bacteria and slime molds, mobile cancer cells may exhibit a suite of collective behaviors and that investigating collective patterns in such devastating cancer systems with an interdisciplinary perspective, including insights gained from other biosystems, has the potential to advance our understanding of the disease and thus may ultimately allow us to therapeutically impact its spatiotemporal dynamics more effectively. The following section briefly describes our concept as to why "swarm-like" behavior should be advantageous for a cancer system and how it may arise.

Paradigms

A central component that underlies our concepts is that cells, also in their diseased state, can function as sender and receiver of chemical (autocrine/paracrine) and biomechanical signals, features supported by a solid body of experimental evidence.⁽¹⁵⁾ Furthermore, the abnormalities present in diseased cells can result in them exhibiting properties reminiscent of those employed by cell aggregates during natural morphogenic processes. Thus collective migration of cells represents a reversion to a type of collective action similar to that evoked in a developmental context. Importantly, however, being out of context of normal morphogenesis and incorporating one, or several, attributes exhibited by less differentiated cells, this aberrant cellular process is similar to, but distinct from, normal developmental mechanisms. The ideas that we outline here also employ key concepts from the study of collective animal behavior, which we propose are also

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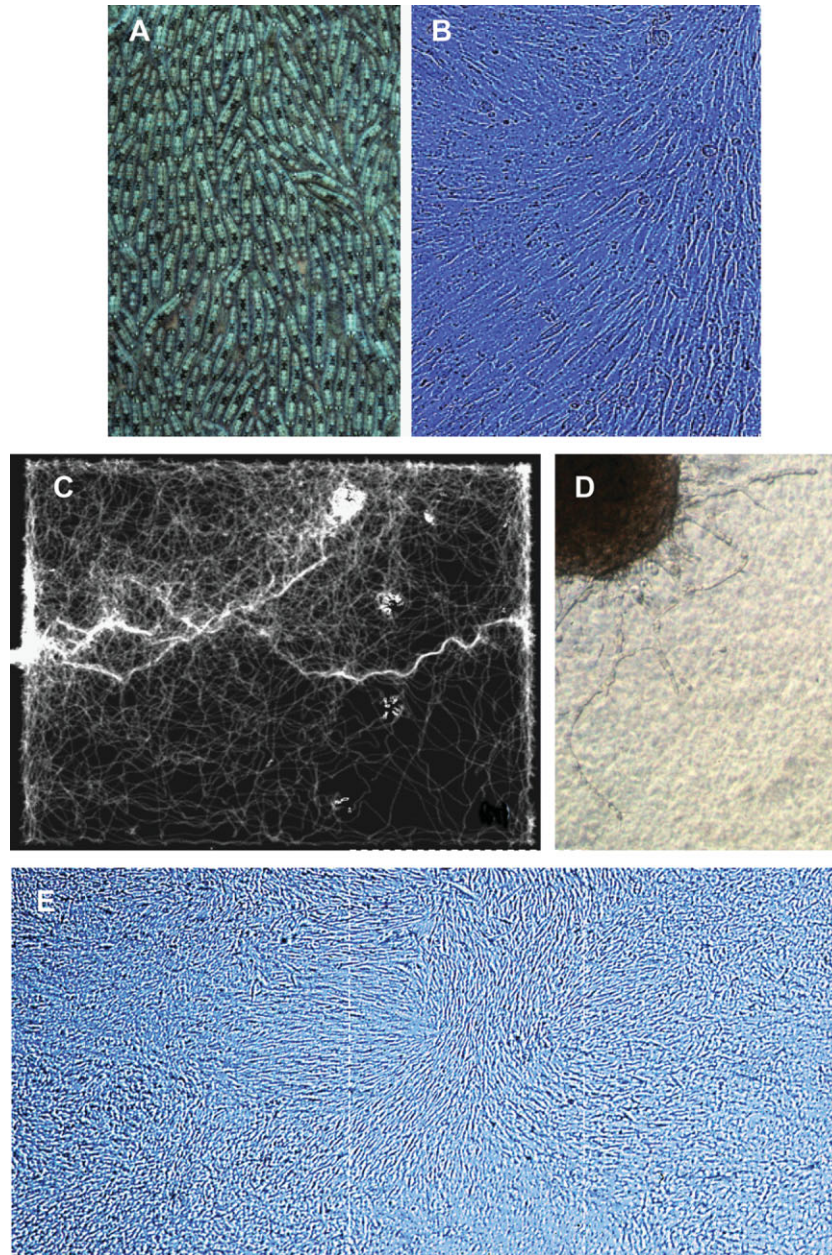


Figure 1. (A) Social caterpillars (image courtesy of Trond Larsen), *cf.* (B) Multicellular patterns that emerge in a confluent human glioma cell culture *in vitro*. Note the resemblance of the two complex patterns, although the characteristic length-scale of the two systems is distinctively different (i.e., a social caterpillar measures several millimeters in length, whereas the diameter of tumor cells is measured in micrometers). (C) Ant trails that originate from the nest (bright spot on the left) to sugar drops (four) deposited on the right (from digital tracking of ants by I. D. C.). (D) Invasive cell branches that originate from a (top left) glioma spheroid *in vitro* (3D extracellular matrix assay using MatrigelTM(⁸), used also as basis for computational cancer modeling works^(70,71)). (E) Microscopic image of a confluent human glioma cell culture *in vitro*. Note the alignment of multiple cancer cells (within the broken lines), as compared to the rest of the cell culture.

highly relevant to understanding the interactions of tumor cell populations. In addition, we argue that similarities among these systems go beyond mere analogy and instead we can begin to make testable predictions about the types, and consequences, of collective behavior within tumors.

Chemical feedback processes and collective migration in tumors

A crucial and ubiquitous form of collective communication in animal groups arises from the interplay of processes that result in amplification and decay of information. Consider, for

example, an ant colony foraging for new resources. Here, individuals can influence each others activity through the deposition of, and response to, chemical pheromones. The chemical acts to modify the environment through which other individuals move, influencing both the direction taken by ants that pass that point in the future and also often attracting (recruiting) ants to a site.⁽¹⁶⁾ Individuals, if capable of depositing pheromone, may reinforce the trail, further influencing the direction taken by subsequent ants, and so on. This social amplification, or positive feedback, can result in exploratory columns of individuals (Fig. 1C). Negative feedback in this system results from the decay (diffusion) of pheromone. If paths are not properly reinforced they fail to elicit a sufficiently strong collective response and foraging columns cannot be maintained. Once trails do form, however, the local aggregation of chemical-producing individuals means that trail persistence can often be maintained with a chemical deposition rate lower than that initially required to define its formation.⁽¹⁷⁾ If positive feedback is high (such as when ants release large quantities of chemical), trails form quickly but are sensitive to initial conditions (random fluctuations). Conversely, if negative feedback dominates, ants will not sufficiently reinforce each others motion and exploration tends to be non-existent or individualistic. Thus, by tuning the rate of reinforcement to that of decay, ant colonies can trade off exploration of new areas with the exploitation of existing resources.⁽¹⁸⁾

The social amplification of collective motion through chemicals means in ants bears similarity to certain processes evident within tumors; processes which emerge as a result of the cells' capability to create and respond to chemicals that also generate collective motion. Consider invasive gliomas, for example (as shown in Figs. 1B and D). These highly malignant brain tumor cells reportedly utilize the amino acid transporter system x_c to import cystine in exchange for the release of relatively large quantities of glutamate.^(19–22) Glutamate, an excitatory neurotransmitter,⁽²³⁾ appears to induce local collective migration of these cancer cells.^(21,24) Consequently, we hypothesize that positive feedback processes also occur here, with glutamate-secreting cell motion being influenced by the inhomogeneities in glutamate concentration (gradients) that the cells, themselves, create. Thus, regions of growing concentration tend to become “reinforced” (autocatalytic behavior), breaking symmetry, and resulting in columns of cells emerging from the main tumor and thus a pattern similar to that shown in Fig. 1D (other potential chemical cues include autocrine production of and paracrine stimulus through transforming growth factor alpha, $TGF\alpha$).⁽¹⁵⁾ In addition to promoting collective cell migration, glutamate production may also act to “clear a path.” That is, by creating a locally inhospitable environment that leads to neuronal death,^(20,21) it can enhance the inflammatory response and may promote peritumoral edema.^(22,24) Thus,

glutamate production acts both to induce collective invasion and to create a more permeable space through which an invasion process can occur that requires less energy dissipation⁽²¹⁾ (a process furthered by the secretion of proteolytic enzymes,⁽²⁵⁾ which already have been linked to collective cancer cell invasion⁽²⁶⁾).

Unlike the case for ants, the positive and negative feedback processes that determine autocatalytic cell migration in tumors are not finely tuned by the requirements of optimal foraging due to the limitations of tumors to adapt primarily by clonal evolution.^(27–29) Nevertheless, an understanding of the onset and maintenance of collective invasion may benefit from considering how these properties can be manipulated, both by internal tumor dynamics and by treatment protocols.

Leadership and collective decision-making in mobile cell aggregates

In some collective tumor invasion processes, it appears that a pro-migratory subset of cells at the leading edge guides the invasion process,^(8,12,30,31) although the extent to which guided cells are completely passive, as argued previously, is unclear. The fairly precise maintenance of cell position with respect to neighbors within the moving group is also demonstrated in models of collective motion in animals where all individuals are capable of active propulsion.^(32,33) Despite this, it is clear that in both developmental processes involving cell migration^(34,35) and tumorigenesis^(8,12,30,31) a relatively small proportion of “trail-blazing” cells can often act to guide others. In some cases this can result in the detachment of cell groups (so-called nests), whereas in others contact is maintained with the primary site.⁽¹²⁾

These processes bear resemblance to leadership behavior investigated in certain animal groups. In some ant species, for example, relatively few “pathfinder ants”⁽³⁶⁾ tend to be the individuals responsible for trail pheromone deposition. Couzin *et al.*⁽³⁷⁾ also demonstrated that only a small proportion of individuals with biased motion (such as cells with directed motion, e.g., those navigating up a chemical gradient) can guide “naive” individuals (those without a preferred direction of travel, or that are incapable of sensing the chemical gradient) without any chemical signaling. If individuals exhibit local attraction (adhesion) to near neighbors, and in addition a tendency to align their direction of travel with such neighbors (caused perhaps by the shear forces generated among motile cells in contact^(38,39)), such leadership can emerge spontaneously. Individuals with a directional bias tend to move to occupy the leading edge of the aggregate regardless of starting positions. Furthermore, it was demonstrated that as the population size increases the proportion of such biased (guiding) individuals required to

induce group migration actually decreases. Therefore, only a very small proportion of actively “invasive” individuals is needed to cause a transition to collective motion of a large body of cells. Importantly, this behavior does not require explicit signaling, individual recognition of cell types nor the production of chemical attractant by leading individuals (however, such physically mediated information transfer is not mutually exclusive from that involving chemical means).

One of the predictions of the model of Couzin *et al.*⁽³⁷⁾ is that, where there is a conflict in the directional preference exhibited by leading individuals, the cell group will either integrate the preferred directions among leading edges (when the angular difference between them is smaller than a critical value; 130° in their model) or collectively select one direction—that associated with the dominant leading edge (such consensus decision-making again results from explicit local interactions). Data already exist that demonstrate that net migration of cancer cell groups in the presence of conflicting leading edges either follow the integrated vector or the dominant leading edge,⁽⁴⁰⁾ and we argue that further analysis of such properties could be very revealing about the underlying mechanism of guided motion in invasive tumor cell aggregates.

Furthermore, a recent investigation of large motile tumor cell sheets both *in vivo* and *in vitro* has revealed that podoplanin, a small mucin-like protein, is up-regulated in the leading edge of invasive structures.⁽⁴¹⁾ We hypothesize that this differentiation of cells within the tumor may relate to the leadership processes we have describe here. What is unclear is whether leading cells change expression after they adopt their position, or whether they become leaders because of the change in expression. In either case the subsequent action of podoplanin acts to promote migration and invasion of cells in a manner reminiscent of the glutamate release in gliomas, as described above. Thus, a similar relationship in terms of positive and negative feedback that we hypothesized previously may also be relevant to a wide range of tumor types, as podoplanin up-regulation is a feature seen in, e.g., squamous cell carcinoma of the lung, skin, cervix, oral cavity, larynx, granulose cell tumors, and a large number of tumors of the central nervous system.⁽⁴¹⁾ Podoplanin is thought to modulate the actin cytoskeleton of cells.⁽⁴¹⁾ Similarly, other pathways that act on cytoskeleton elements, as well as those involved in cell–cell (and cell–matrix) interactions (such as integrins⁽³¹⁾) should be expected to influence the collective dynamics of invasion.⁽⁴²⁾

Density-dependent transitions to collective motion

As discussed above, the transition from quasi-stable aggregates to those that exhibit collective migration can occur as a result of the release of attractant and/or motility-

inducing chemicals from even a relatively small proportion of cells. This appears to result frequently in the formation of tendril or cone-like extensions,⁽⁴¹⁾ strands, columns or the dispersal of rafts, or “nests” of cells.⁽³⁰⁾ As stated above, the role of cell–cell adhesion to such processes is highly important and modifications to pathways influencing adhesion can have a dramatic effect on the cohesive nature of such migratory patterns.

Cell adhesion molecules (surface glycoproteins) also play an important role in coordinating the spatial distribution of motile cells within aggregates. Due to principles of energy minimization,^(38,43) cells with similar surface chemistry tend to become spontaneously assorted within aggregates. This is seen with differentiated cells can accumulate with others from their respective tissue after being mixed in culture.^(44–47) Similar processes can explain cell sorting in embryogenesis and that within cellular swarms.⁽⁴⁸⁾ Computer models in which cell surface fluctuations and differential adhesion among cell types are simulated (often using what is known as the extended Potts model⁽³⁸⁾) replicate such processes well and, furthermore, demonstrate how adhesion among motile cells causes shear stresses that tend to align the local direction of travel of individuals.⁽³⁹⁾ In fact, the traction exerted by extracellular matrix remodeling cells can lead to so-called “contact guidance,” i.e., a stress–strain response alignment of cells along reoriented fibrils within the microenvironment.⁽⁴⁹⁾ The result is a biased migration seen not only in fibroblasts⁽⁵⁰⁾ but also in, for example, highly invasive MV3 melanoma cancer cells, where it reportedly induced the recruitment of proximal cancer cells to migrate along preexisting routes of matrix compaction, reorganized by previous passenger cells.⁽⁵¹⁾

Recent studies of collective motion in animal groups have demonstrated both theoretically,⁽⁵⁾ and empirically,⁽³⁾ that such local alignment can spread rapidly across populations if individuals are motile, resulting in the generation of order (directed motion) across length-scales much greater than the individual interaction range (see also Fig. 1). Importantly, these studies reveal that a sudden, and spontaneous, transition exists between disorder (where individual motion within the aggregate appears relatively erratic and uncorrelated with others) and order (where individual motion is correlated over long distances) as the density of interacting elements is increased. In mass-migrating insects (locusts),⁽³⁾ and also keratinocytes,⁽⁵²⁾ this transition has been experimentally validated. Furthermore, statistical mechanics has revealed that, when such large interacting systems change population-level properties in this way (a so-called “phase transition,” analogous to sudden transitions in physical systems such as from a liquid to a gas), the change in collective behavior is independent of the details of the interacting components,⁽⁵³⁾ a feature known as “universality.”

We hypothesize that similar density-dependent transitions are likely to occur within aggregates of motile tumor cells. Analysis of cell orientation in mammalian skin has already suggested that such mechanisms are plausible.⁽⁵⁴⁾ Thus, there need not necessarily be a “signal” to determine the onset of migration. Rather, it could occur spontaneously when the density of cells exceeds a critical amount. As noted before, such transitions may occur in concert with other mechanisms of orientation—our hypotheses are not mutually exclusive, but they are testable. Correlates of density and alignment made from tumor samples could be highly informative, and density manipulations *in vitro* may allow for specific replication of the spontaneous density-dependent transition to mass migration. Furthermore, for a given density of cells, treatments inhibiting, or enhancing, the mutual shear forces expressed by cell bodies (and consequently their ability to align with neighbors with whom they are in contact) should be expected to also change populations from disorder to ordered states.

Benefits of long-range information transfer

The ability for cells to interact locally and yet to form cohesive moving sheets may have important implications to navigational and resource-finding properties of tumor cells. As has been suggested for animal groups,^(55,56) coherent interactions among cells may allow them to form an integrated self-organizing array of “sensors” capable of damping local fluctuations and thus reliably detecting, and responding to, weak long-range gradients. As such, a swarm-like search formation may reduce randomness and metabolic cost for the individual, while increasing the probability of locating resources for the collective.

Cell–cell information processing may also act to alert bystanders about a supra-regional microenvironment without them physically having to move closer to the sites to collect this information. This strategy reduces per cell motility, hence minimizes energy dissipation of the individual while optimizing the yield for the entire population. Aggregation by cells when migrating may also confer other benefits. Similar to the schooling by prey fish making it harder for a predator to locate them, a tight multi-cell formation exposes less individual cell surface to potentially adverse environmental conditions, such as anti-cancerous immune cells and chemotherapeutic agents.⁽⁵⁷⁾ Conversely, in the presence of abundant nutrients, a swarm-like formation can quickly expand the tumor system’s invasive surface toward the source. The result is an increased “surface-to-volume” ratio within the mobile cell population that maximizes nutrient uptake per cell. At the same time, despite angiogenesis, this very same relationship is poised to grow negatively for the main tumor mass (which in turn is thought to be responsible for the onset of central necrosis, and could be a trigger for the onset of invasion to begin with).

Conflict, cooperation, and the onset of swarm-like behavior

Lastly, one could hypothesize that swarm-like behavior is prevalent when the microenvironment is not very permissive, and cell–cell “cooperation”⁽²⁷⁾ is key to ensure survival of a more heterogeneous cell population. Indeed, other cellular swarms, such as those formed by bacteria and slime mold (*Dictyostelium discoideum*) cells, respond to adverse and variable environmental conditions by employing sophisticated cooperative behavior involving chemical signaling and collective migration.^(1,48,58) In such systems, chemical communication among cells involving low molecular mass molecules allows monitoring of population density and a switch to a collective response once a quorum density is reached. “Quorum-regulated swarming,” which itself involves chemical signaling, is thought to enhance motility of cells once local resources are insufficient to support colony growth.^(1,59) Eventually, however, longer-term stress can result in “competition” becoming a dominant feature. Here, selective advantage leads to the emergence of dominant clones best fit for the present conditions.⁽⁵⁹⁾ As in tumors, cells inside growing bacterial swarms may die, resulting in a proliferating edge and necrotic core.⁽⁶⁰⁾

Sudden bursts of growth at the edge, called “sectoring,” also occur under harsh conditions in bacterial swarms. These sudden bursts of growth give rise to phenotypically recognizable sectors and appear to arise from a single mutant ancestral cell superior in motility or growth rate.⁽⁶⁰⁾ This strategy can facilitate long-range food finding, but comes at a cost: due to the high energetic requirements of this growth the cells are susceptible to both extreme stress and antibiotic therapy. Under such conditions slow-growing, metabolically quiescent cells are most likely to survive.

Thus, conceivably, collective behavior, or more generally, the emergence of cooperation within tumors may be bound by an upper and lower threshold of microenvironmental conditions. Since, the distribution of these microenvironmental conditions is hardly homogeneous, in reality a tumor system likely displays “heterogeneous” patterns throughout, i.e., at a given point in time, some sections show collective behavior while others do not. This appears to be supported by Fig. 1E, which shows a confluent glioma cell monolayer *in vitro*

Unlike social insect, bacterial and slime mold swarms, collective strategies of cancer cells cannot evolve the same level of sophisticated adaptive response (since a “successful” strategy likely kills the host and as such the information does not pass on to another generation). Rather, the tumors evolutionary process is limited to clonal evolution within the relatively short context, in evolutionary terms, of the host body. However, as we have described above, driven by competition and cooperation, populations of cancer cells may be able to tap into suites of complex and coordinated collective behavior

that have evolved for other purposes within organisms such as development and immune response.^(9,30,42,46,54,61,62)

Conclusions

If the mobile fraction of a cancer system is indeed capable of showing such swarm-like behavior, we argue that it would not be reasonable to try to randomly target single invasive cells. The characteristic of such collective behavior is that it can compensate the loss of a fair number of its constituents without losing its overall appearance or “performance.” In the case of a highly proliferative tumor system, one would in addition have to deal with the continuous production of these individuals into the swarm. This suggests therapeutic strategies with different degrees of feasibility at present. These are summarized below:

There should be a “critical density” of mobile cancer cells below which no such collective behavior can exist and thus the cell–cell signal intensity would drop below the lower detection threshold. If this is so, then, in theory, broadly targeting the entire mobile cell population could be effective. In reality, however, mobile cells currently cannot be imaged clinically, much less targeted, and are also, at least for the case of brain cancer,⁽⁶³⁾ thought to be non-proliferative. The latter means that even if one could eventually detect and target single invasive cells, they cannot be treated with conventional anti-proliferative treatments such as radiotherapy or chemotherapy. Regardless, targeting an extensive tissue area (in an attempt of killing a significant number of invasive cells) will inevitably have side effects for the healthy tissue they invade, rendering it prohibitively “costly” for the patient.

Another, conceptually more appealing strategy would be geared toward “interrupting” the cell swarm’s information process. That is, if cell–cell communication can be stopped or at least severely hampered therapeutically, arguably the system overall would slow down. That approach could be particularly promising if (parts of) the cell–cell information network turned out to be more connected than others, and thus present higher “value” targets. Furthermore, if the phenotypic abilities of the trail-blazing cells indeed marked a distinctively aggressive subclonal cancer cell population, rather than being the result of dynamic gene expression changes (see “*Leadership and collective decision-making in mobile cell aggregates*” above), one should gain therapeutically by targeting these guiding “tip” cells selectively. Intriguingly, inhibiting the afore-mentioned x_c pathway and limiting cystine uptake reduces cellular invasion and tumor growth.⁽¹⁹⁾ However, aside from the considerable technical challenges related to first detecting and then genetically engineering a subset of tumor cells on site, the outcome appears dubious (even if technically achievable at some point). As we stressed in the previous section, conceivably,

collective and competitive behavior can occur side-by-side as it may simply depend on microenvironmental conditions and the reactions they trigger. That being said, a therapeutic decoupling of mobile tumor cells from the swarm may very well lead them to behave competitively and leave them largely uncontrolled. Indeed, the experimental literature lists plenty of evidence that reducing cell–cell communication, e.g., by targeting gap-junctions, yields a more aggressive tumor system.^(64,65)

More promising than targeting or fragmenting it, should be exploiting the capability of processing information by the invasive cell swarm and thus taking over its directional control. Specifically, one can envision “redirecting” the mobile cells by for instance implanting small, growth factor-releasing, biodegradable polymer wavers into the surrounding tissue, hence utilizing recent advances made in chemotherapeutic delivery.^(66,67) It would be important, through proper labeling, to ensure that the wavers can be detected with advanced imaging modalities. The wavers sole task would be to serve as a local chemotactic source to attract tumor cells and, thus, by means of cell–cell communication, to lead the cancer cell swarm away from physiologically critical or surgically not readily accessible tissue areas. In a second therapeutic step, these wavers together with their bystander tumor cells could then be targeted specifically—hopefully then with a lesser concern for the parenchyma. Limitations of this innovative strategy include biocompatibility of implanted materials and the concern that utilizing, if not inducing, collective cancer cell mobility without taming the cells’ matrix-degrading enzymes could even exacerbate tissue damage. While the technical challenges will undoubtedly be considerable, the goal would be in fact not as much cure, but a better “controlled” tumor system.

Of course, much work has to be done experimentally before a move towards clinics can even be considered. To start with, the anecdotal reports of such *in vitro* behavior will have to be investigated methodically with non-invasive imaging techniques to analyze the time series, and the mechanisms by which cells stay in contact have to be reevaluated from that perspective. If our paradigm still holds, the question arises of how much, if any, relevance such *in vitro* findings bear for the *in vivo* situation. In real tissues, the spatio-temporal distribution dynamics of any diffusive signal will be quite different. Adding the comparably very low per cell secretion rates of any of these messenger cues or paracrine growth factors, one has to wonder if the considerable structural constraints would even allow these cells to display any such behavior. Under these heterogeneous 3D conditions, “biomechanical” signaling may play a more prominent role. Partly, based on the concept that proteolysis impacts microenvironmental biomechanics, it could be argued that collective behavior is also involved in the “beaten-path” phenomenon that has invasive cells seemingly following one

another along the path of “least resistance, most permission, and highest attraction,” which in turn has been proposed to help generate tumor cell branching patterns seen in 3D matrix gel environments (see refs. ^(8,68,69) and Fig. 1D). Similar mechanisms also exist in animal groups where local modification of the environment, by deposition of chemicals such as ants’ chemical trails and/or physical means (by locally decreasing resistance to motion) results in path formation. It should be noted, however, that these processes are not mutually exclusive and considerable functional complexity can arise in interacting systems even when interactions are explicitly local.⁽¹⁾

In summary, if cancer indeed represents a complex dynamic biosystem, it seems odd to argue that their mobile cell population should not be capable of exhibiting collective behavior similar to that of other biological systems. Therefore, investigating such swarm-like behavior in more detail is important from a cancer research and, if confirmed, even more so from a clinical oncology perspective.

Acknowledgments: This work has been supported in part by NIH grant CA 113004 and by the Harvard-MIT (HST) Athinoula A. Martinos Center for Biomedical Imaging and the Department of Radiology at Massachusetts General Hospital. T. S. D. thanks Dr. Zhihui Wang (Massachusetts General Hospital) for help with formatting the manuscript. I. D. C. is indebted to Nicola Grimwood for her contribution and gratefully acknowledges support from a Searle Scholar Award and also a DARPA grant HR001-05-1-0057 to Princeton University.

References

1. Camazine, S., Deneubourg, J.-L., Franks, N. R., Sneyd, J., Theraulaz, G. and Bonabeau, E., Self-organization in biological systems. Princeton, Princeton University Press, 2002.
2. Deisboeck, T. S. and Kresh, J. Y., Complex systems science in biomedicine. New York, Springer, 2006.
3. Buhl, J., Sumpter, D. J., Couzin, I. D., Hale, J. J., Despland, E., et al. From disorder to order in marching locusts. *Science* 2006. **312**(5778):1402–1406.
4. Levin, S. A., The problem of pattern and scale in ecology: the Robert H. MacArthur award lecture. *Ecology* 1992. **73**(6):1943–1967.
5. Vicsek, T., Czirok, A., Ben-Jacob, E., Cohen, I. I. and Shochet, O., Novel type of phase transition in a system of self-driven particles. *Phys Rev Lett* 1995. **75**(6):1226–1229.
6. Yeomans, J. M., Statistical mechanics of phase transitions. Oxford, Oxford University Press, 1992.
7. Coffey, D. S., Self-organization, complexity and chaos: the new biology for medicine. *Nat Med* 1998. **4**(8):882–885.
8. Deisboeck, T. S., Berens, M. E., Kansal, A. R., Torquato, S., Stemmer-Rachamimov, A. O. and Chiocca E. A., Pattern of self-organization in tumour systems: complex growth dynamics in a novel brain tumour spheroid model. *Cell Prolif* 2001. **34**(2):115–134.
9. Kenny, P. A., Nelson, C. M. and Bissell, M. J., The ecology of tumors. *The Scientist* 2006. **20**(4):31–35.
10. Hanahan, D. and Weinberg, R. A., The hallmarks of cancer. *Cell* 2000. **100**(1):57–70.
11. Fidler, I. J., The pathogenesis of cancer metastasis: the ‘seed and soil’ hypothesis revisited. *Nat Rev Cancer* 2003. **3**(6):453–458.
12. Friedl, P. and Wolf, K., Tumour-cell invasion and migration: diversity and escape mechanisms. *Nat Rev Cancer* 2003. **3**(5):362–374.
13. Bidard, F. C., Pierga, J. Y., Vincent-Salomon, A. J. and Poupon, M. F., A “class action” against the microenvironment: do cancer cells cooperate in metastasis? *Cancer Metastasis Rev* 2008. **27**(1):5–10.
14. Ewald, A. J., Brenot, A., Duong, M., Chan, B. S. J. and Werb, Z., Collective epithelial migration and cell rearrangements drive mammary branching morphogenesis. *Dev Cell* 2008. **14**(4):570–581.
15. Ekstrand, A. J., James, C. D., Cavenee, W. K., Seliger, B., Pettersson, R. F. and Collins V. P., Genes for epidermal growth factor receptor, transforming growth factor alpha, and epidermal growth factor and their expression in human gliomas *in vivo*. *Cancer Res* 1991. **51**(8):2164–2172.
16. Hölldobler, B. and Wilson, E. O., The ants. Boston, Harvard University Press, 1990.
17. Beekman, M., Sumpter, D. J. and Ratnieks, F. L., Phase transition between disordered and ordered foraging in Pharaoh’s ants. *Proc Natl Acad Sci USA* 2001. **98**(17):9703–9706.
18. Deneubourg, J.-L., Aron, S., Goss, S., Pasteels, J. M. and Deurincq, G., Random behaviour, amplification processes and number of participants: how they contribute to the foraging properties of ants. *Physica D* 1986. **2**(1–3):176–186.
19. Chung, W. J., Lyons, S. A., Nelson, G. M., Hamza, H., Gladson, C. L., et al. Inhibition of cystine uptake disrupts the growth of primary brain tumors. *J Neurosci* 2005. **25**(31):7101–7110.
20. Domerq, M., Sanchez-Gomez, M. V., Sherwin, C., Etxebarria, E., Fern, R. and Matute C., System xc- and glutamate transporter inhibition mediates microglial toxicity to oligodendrocytes. *J Immunol* 2007. **178**(10):6549–6556.
21. Sontheimer, H., Malignant gliomas: perverting glutamate and ion homeostasis for selective advantage. *Trends Neurosci* 2003. **26**(10):543–549.
22. Takano, T., Lin, J. H., Arcuino, G., Gao, Q. J., Yang, J. and Nedergaard M., Glutamate release promotes growth of malignant gliomas. *Nat Med* 2001. **7**(9):1010–1015.
23. Fellin, T., Pascual, O., Gobbo, S., Pozzan, T., Haydon, P. G. and Carmignoto G., Neuronal synchrony mediated by astrocytic glutamate through activation of extrasynaptic NMDA receptors. *Neuron* 2004. **43**(5):729–743.
24. Lyons, S. A., Chung, W. J., Weaver, A. K., Ogunrinu, T. and Sontheimer, H., Autocrine glutamate signaling promotes glioma cell invasion. *Cancer Res* 2007. **67**(19):9463–9471.
25. Nabeshima, K., Inoue, T., Shima, Y., Okada, Y., Itoh, Y., et al. Front-cell-specific expression of membrane-type 1 matrix metalloproteinase and gelatinase A during cohort migration of colon carcinoma cells induced by hepatocyte growth factor/scatter factor. *Cancer Res* 2000. **60**(13):3364–3369.
26. Friedl, P. and Wolf, K., Tube travel: the role of proteases in individual and collective cancer cell invasion. *Cancer Res* 2008. **68**(18):7247–7249.
27. Axelrod, R., Axelrod, D. E. and Pienta, K. J., Evolution of cooperation among tumor cells. *Proc Natl Acad Sci USA* 2006. **103**(36):13474–13479.
28. Merlo, L. M., Pepper, J. W., Reid, B. J. and Maley, C. C., Cancer as an evolutionary and ecological process. *Nat Rev Cancer* 2006. **6**(12):924–935.
29. Nowell, P. C., The clonal evolution of tumor cell populations. *Science* 1976. **194**(4260):23–28.
30. Friedl, P., Prespecification and plasticity: shifting mechanisms of cell migration. *Curr Opin Cell Biol* 2004. **16**(1):14–23.
31. Hegerfeldt, Y., Tusch, M., Brocker, E. B. and Friedl, P., Collective cell movement in primary melanoma explants: plasticity of cell–cell interaction, beta1-integrin function, and migration strategies. *Cancer Res* 2002. **62**(7):2125–2130.
32. Couzin, I. D., Krause, J., James, R., Ruxton, G. D. and Franks, N. R., Collective memory and spatial sorting in animal groups. *J Theor Biol* 2002. **218**(1):1–11.
33. Gregoire, G., Chate, H. and Tu, Y., Moving and staying together without a leader. *Physica D* 2003. **181**: 157–170.

34. Haas, P. and Gilmour, D., Chemokine signaling mediates self-organizing tissue migration in the zebrafish lateral line. *Dev Cell* 2006. **10**(5):673–680.
35. Insall, R. H. and Jones, G. E., Moving matters: signals and mechanisms in directed cell migration. *Nat Cell Biol* 2006. **8**(8):776–779.
36. Jackson, D. E., Martin, S. J., Ratneiks, F. L. W. and Holcombe, M., Spatial and temporal variation in pheromone composition of ant foraging trails. *Behav Ecol* 2007. **18**: 444–450.
37. Couzin, I. D., Krause, J., Franks, N. R. and Levin, S. A., Effective leadership and decision-making in animal groups on the move. *Nature* 2005. **433**(7025):513–516.
38. Glazier, J. A. and Graner, F., Simulation of the differential adhesion driven rearrangement of biological cells. *Phys Rev E Stat Phys Plasmas Fluids Relat Interdiscip Topics* 1993. **47**(3):2128–2154.
39. Rappel, W.-J., Nicol, A., Sarkissian, A., Levine, H. and Loomis, W. F., Self-organized vortex state in two-dimensional *Dictyostelium* dynamics. *Phys Rev Lett* 1999. **83**: 1247–1250.
40. Kolega, J., The movement of cell clusters *in vitro*: morphology and directionality. *J Cell Sci* 1981. **49**: 15–32.
41. Wicki, A. and Christofori, G., The potential role of podoplanin in tumour invasion. *Br J Cancer* 2007. **96**(1):1–5.
42. Mouneimne, G. and Brugge, J. S., Tensins: a new switch in cell migration. *Dev Cell* 2007. **13**(3):317–319.
43. Beysens, D. A., Forgacs, G. and Glazier, J. A., Cell sorting is analogous to phase ordering in fluids. *Proc Natl Acad Sci USA* 2000. **97**(17):9467–9471.
44. Moscona, A., The development *in Vitro* of chimeric aggregates of dissociated embryonic chick and mouse cells. *Proc Natl Acad Sci USA* 1957. **43**(1):184–194.
45. Steinberg, M. S., Reconstruction of tissues by dissociated cells. Some morphogenetic tissue movements and the sorting out of embryonic cells may have a common explanation. *Science* 1963. **141**: 401–408.
46. Townes, P. L. and Holtfreter, J., Directed movements and selective adhesion of embryonic amphibian cells. *J Exp Zool* 1955. **128**: 53–120.
47. Trinkaus, J. P. and Groves, P. W., Differentiation in culture of mixed aggregates of dissociated tissue cells. *Proc Natl Acad Sci USA* 1955. **41**(10):787–795.
48. Maree, A. F. and Hogeweg, P., How amoeboids self-organize into a fruiting body: multicellular coordination in *Dictyostelium discoideum*. *Proc Natl Acad Sci USA* 2001. **98**(7):3879–3883.
49. Tranquillo, R. T., Self-organization of tissue-equivalents: the nature and role of contact guidance. *Biochem Soc Symp* 1999. **65**: 27–42.
50. Dickinson, R. B., Guido, S. and Tranquillo, R. T., Biased cell migration of fibroblasts exhibiting contact guidance in oriented collagen gels. *Ann Biomed Eng* 1994. **22**(4):342–356.
51. Friedl, P., Maaser, K., Klein, C. E., Niggemann, B., Krohne, G. and Zänker, K. S., Migration of highly aggressive MV3 melanoma cells in 3-dimensional collagen lattices results in local matrix reorganization and shedding of alpha2 and beta1 integrins and CD44. *Cancer Res* 1997. **57**(10):2061–2070.
52. Szabo, B., Szollosi, G. J., Gonci, B., Juranyi, Z., Selmeczi, D., *et al.* Phase transition in the collective migration of tissue cells: experiment and model. *Phys Rev E Stat Nonlin Soft Matter Phys* 2006. **74**(6):061908.
53. Stanley, H. E., Scaling, universality and renormalization: three pillars of modern critical phenomena. *Rev Mod Phys* 1999. **71**(2):S358–366.
54. Wang, Y., Badea, T. and Nathans, J., Order from disorder: Self-organization in mammalian hair patterning. *Proc Natl Acad Sci USA* 2006. **103**(52):19800–19805.
55. Couzin, I., Collective minds. *Nature* 2007. **445**(7129):715.
56. Grünbaum, D., Schooling as a strategy for taxis in a noisy environment. *Evol. Ecol.* 1998. **12**(5):503–522.
57. Christiansen, J. J. and Rajasekaran, A. K., Reassessing epithelial to mesenchymal transition as a prerequisite for carcinoma invasion and metastasis. *Cancer Res* 2006. **66**(17):8319–8326.
58. Gilbert, O. M., Foster, K. R., Mehdiabadi, N. J., Strassmann, J. E. and Queller, D. C., High relatedness maintains multicellular cooperation in a social amoeba by controlling cheater mutants. *Proc Natl Acad Sci USA* 2007. **104**(21):8913–8917.
59. Ben-Jacob, E. and Aharonov, Y. Y. S., Bacteria harnessing complexity. *Biofilms* 2004. **1**: 239–263.
60. Ron, I. G., Golding, I., Lifshitz-Mercer, B. and Ben-Jacob, E., Bursts of sectors in expanding bacterial colonies as a possible model for tumor growth and metastases. *Physica A* 2003. **320**: 485–496.
61. Hanisch, U. K. and Kettenmann, H., Microglia: active sensor and versatile effector cells in the normal and pathologic brain. *Nat Neurosci* 2007. **10**(11):1387–1394.
62. Huang, S., Brangwynne, C. P., Parker, K. K. and Ingber, D. E., Symmetry-breaking in mammalian cell cohort migration during tissue pattern formation: role of random-walk persistence. *Cell Motil Cytoskeleton* 2005. **61**(4):201–213.
63. Giese, A., Loo, M. A., Tran, N., Haskett, D., Coons, S. W. and Berens, M. E., Dichotomy of astrocytoma migration and proliferation. *Int J Cancer* 1996. **67**(2):275–282.
64. McDonough, W. S., Johansson, A., Joffe, H., Giese, A. and Berens, M. E., Gap junction intercellular communication in gliomas is inversely related to cell motility. *Int J Dev Neurosci* 1999. **17**(5–6):601–611.
65. Oliveira, R., Christov, C., Guillamo, J. S., de Bouard, S., Palfi, S., *et al.* Contribution of gap junctional communication between tumor cells and astroglia to the invasion of the brain parenchyma by human glioblastomas. *BMC Cell Biol* 2005. **6**(1):7.
66. Fleming, A. B. and Saltzman, W. M., Pharmacokinetics of the carmustine implant. *Clin Pharmacokinet* 2002. **41**(6):403–419.
67. Westphal, M., Ram, Z., Riddle, V., Hilt, D. and Bortey, E., Gliadel wafer in initial surgery for malignant glioma: long-term follow-up of a multicenter controlled trial. *Acta Neurochir (Wien)* 2006. **148**(3):269–275; discussion 275.
68. Gordon, V. D., Valentine, M. T., Gardel, M. L., Andor-Ardo, D., Dennison, S., *et al.* Measuring the mechanical stress induced by an expanding multicellular tumor system: a case study. *Exp Cell Res* 2003. **289**(1):58–566.
69. Kaufman, L. J., Brangwynne, C. P., Kasza, K. E., Filippidi, E., Gordon, V. D., *et al.* Glioma expansion in collagen I matrices: analyzing collagen concentration-dependent growth and motility patterns. *Biophys J* 2005. **89**(1):635–650.
70. Habib, S., Molina-Paris, C. and Deisboeck, T. S., Complex dynamics of tumors: modeling an emerging brain tumor system with coupled reaction-diffusion equations. *Physica A* 2003. **327**: 501–524.
71. Sander, L. M. and Deisboeck, T. S., Growth patterns of microscopic brain tumors. *Phys Rev E Stat Nonlin Soft Matter Phys* 2002. **66**(5 Pt 1):051901.